



Aflatoxin M₁ in Pasteurized Milk in Babol city, Mazandaran Province, Iran

SAA Sefidgar¹, M Mirzae¹, M Assmar², *SR Naddaf²

¹Dept. of Medical Mycology, School of Medicine, Babol University of Medical Sciences, Babol, Iran

²Dept. of Parasitology, Pasteur Institute of Iran, Tehran, Iran

(Received 29 Jun 2010; accepted 9 Jan 2011)

Abstract

Background: Aflatoxin M₁ (AFM₁) is the metabolite of aflatoxin B₁ (AFB₁) and is found in milk when lactating animals are fed with contaminated feedstuff. The presence of AFM₁ in milk, pose a major risk for humans especially kids as it can have immunosuppressive, mutagenic, teratogenic and carcinogenic effects. The present study is aimed to investigate the occurrence of AFM₁ in subsidized pasteurized milk in Babol, Mazandaran Province, Iran.

Methods: Some 72 pasteurized milk packages were collected from supermarkets in various districts of city during January to March 2006. Milk samples were centrifuged and amounts of 100 µl of skimmed milk were tested for AFM₁ contamination by competitive ELISA.

Results: All the samples (100%) exhibited contamination with AFM₁. The contamination levels means in January, February, and March were 227.85, 229.64, and 233.1ng/l, respectively. The amount of AFM₁ in all the samples were above 50ng/l, the threshold set by the European community regulations.

Conclusion: Monitoring of AFM₁ level should be part of quality control procedures in dairy factories, particularly the ones providing infant's milk. Production of safer and healthier milk and other dairy products with minimum AFM₁ level can be achieved by adopting prophylactic measures including control of humidity and water content of feedstuff, which favors mould production.

Keywords: Aflatoxin M₁, ELISA, Pasteurized milk, Iran

Introduction

Aflatoxins are secondary toxic metabolites and found in most plant products including peanuts, copra, soya, maize, rice and wheat. Aflatoxins B₁, B₂, G₁, and G₂ are produced by special strains of *Aspergillus flavus*, and *A. parasiticus*. The species *A. flavus* merely produces AFB, while other species produce both AFB and AFG(1). Aflatoxins are produced during growth, harvesting and storage course (2). When animals consume AFB₁ contaminated feedstuff, the toxin is metabolized in the liver and excreted as AFM₁ via milk, urination, and feces (3, 4). AFM₁ is bound to milk proteins, especially casein, which leads to its presence in dairy products (2). Aflatoxins are acute toxic compounds, and have shown to be immunosuppressive, mutagen, teratogen and carcinogen

(5). International Agency for Research on Cancer (IARC), WHO introduced Aflatoxins B₁ and M₁ as primary and secondary groups of carcinogenic compounds, respectively (6). The main target organ for toxicity and carcinogenicity is liver. Although mutagenic and carcinogenic level of AFM₁ is lower than AFB₁, its geotaxis activity is known to be much higher (7). AFM₁ is resistant to heat and is not degraded during pasteurization process (8). Food standards for AFM₁ level varies in different countries. According to European community and Codex Alimentary the maximum level for AFM₁ should not exceed 50 ng/kg in raw milk and processed dairy products (4). In Australia and Switzerland AFM₁ levels in children food are not allowed to be more than 10 ng/kg (9). In order to establish a limit for AFM₁

in milk and other dairy products in Iran, preliminary studies are needed to examine its occurrence and level in different areas of Iran.

This study is aimed to investigate the amounts of AFM₁ in subsidized pasteurized milk in Babol, Mazandaran Province, Iran.

Material and Methods

During the winter 2006, some 72 pasteurized milk packages were collected from supermarkets in various districts of Babol and transferred to Department of Medical Mycology, Babol University of Medical Sciences. Milk samples were centrifuged at 3000g for 10 min at 2-8 °C and skimmed milk was directly used for quantitative analysis of AFM₁ by competitive ELISA, using AFLA M₁ Kit- code MA418 (Tecna, Trista, Italy). Simply, one hundred µl of prepared samples and standard solutions were added into each well, already coated with anti-aflatoxin M₁ antibodies by the manufacturer, and incubated at 20-25° C for 45 min. The plates were washed four times by washing buffer, 100 µl enzymes conjugate solution was

added into each well and plates were incubated at 20-25° C for 15 min. The plates were washed as before and 100 µl of developing solution was added into each well and incubated at 20-25° C in a dark place for 15 min. The reaction was stopped by stop solution and absorbance was measured at 450 nm by an ELISA reader within 60 min.

Results

A total of 72 pasteurized milk samples were analyzed with the competitive ELISA. All the samples (100%) were found contaminated with AFM₁. The contamination means in January, February and March were 227.85, 229.64, and 233.1 ng/l, respectively. The highest contamination was observed in March and the lowest in January. AFM₁ contamination ranged from 178.8 to 253.5 ng/l (mean value 230.2), which shows that the contamination level in all samples (100%) exceeded the European community regulations (50 ng/l). There was no significant relationship between AFM₁ contamination level and different months of winter ($P \leq 0.05$) (Table 1, 2).

Table 1: The AFM₁ contamination levels in milk samples during winter 2006 in Babol, Mazandaran Province, Iran

AFM ₁ concentration ng/l	>250	201-250	151-200	51-150	<50
Number of assayed milk samples	4	62	6	0	0
%	5.5	86	8.5	0	0

Table 2: Number and mean values of AFM₁ positive milk samples during winter 2006 in Babol, Mazandaran Province, Iran

Months	Number of Samples tested	Contaminated samples > 50 ng/l, (%)	Mean±SE
Jan.	24	24 (100)	227.85±4.98
Feb.	24	24(100)	229.64±4.19
Mar.	24	24(100)	233.1±4.07
Total	72	72(100)	230.2±1.89

Discussion

The occurrence of Aflatoxin in food is a serious global health problem, particularly in developing countries. Aflatoxins are well documented as cancer potency factors as 4.6-28.2% of annual hepatocarcinoma cases worldwide are caused by these

toxins (10). The presence of AFM₁ in milk and other dairy products is a concern throughout the world and many countries have set threshold limits for milk used by adults and infants. Despite a considerable progress in food industry in Iran during the past two decades, a few reliable data

is available on contamination levels of milk and other dairy products with AFM₁. The earliest study in Iran dates back to 1982, in which 52 liquid milk samples from Tehran showed to be contaminated with AFM₁, ranging from 23 to 3000 µl/l (11). Also, in another study on pasteurized milk samples from Tehran, AFM₁ contamination showed to be above the threshold (50ng/kg) set by European Union (11). However, in two separate studies in Sarab and Shiraz on 111 and 624 milk samples, AFM₁ contamination level was below the maximum tolerance limit of European Union (12). A recent study in Tehran showed that the amount of AFM₁ in 78% of liquid milk samples was higher than the maximum tolerance limit accepted by European Union. However, a validated High Performance Liquid Chromatography (HPLC) analysis of milk samples from five different regions in Iran (Gorgan, Hamadan, Rasht, Shiraz and Tehran) showed that the AFM₁ contamination levels were much lower than those obtained by quantitative ELISA (13). Our results in Babol showed that AFM₁ contamination levels in 100% of milk samples were about 4 folds more than European Union standard. The difference in AFM₁ contamination levels may be attributed to different methods of evaluation and seasonal variations. Thus, adopting a reliable and uniform standard method for measurement of AFM₁ contamination in milk and other dairy products is a crucial step. Monitoring AFM₁ levels should be part of quality control procedures in dairy factories, particularly the ones providing infant's milk, as young animals were found to be more susceptible to aflatoxins than adults (14). An accurate knowledge on the seasonal variation of AFM₁ contamination in milk from different geographical areas in Iran can provide valuable information on factors affecting AFB₁ production, the primary aflatoxin that is metabolized to AFM₁. The amount of AFB₁ in animal feed can be minimized by taking care of cultural phases, including harvest and storage practices, that present critical points for fungal growth and mycotoxin production (2). If reduction of AFM₁ concentration in feedstuff is not possible in some areas, addition of aflatoxin adsorbents to

the contaminated feedstuffs can be considered as an alternative method (2).

Ethical Considerations

Ethical issues including plagiarism, informed consent, misconduct, data fabrication and/or falsification, double publication and/or submission, redundancy, etc. have been completely observed by the authors.

Acknowledgements

We thank our colleagues in Department of Medical Mycology, School of Medicine, Babol University of Medical Sciences for their helpful comments and supplying us with some reagents. This research was financially supported by Babol university of Medical sciences and Sepid private Medical Laboratory, Babol. The authors declare that they have no conflict of interest.

References

1. D'Mello JPF, MacDonald AMC (1997). Mycotoxins. *Animal Feed Sci Tech*, 69: 155-66.
2. Prandini A, Tansini G, Sigolo S, Filippi L, Laporta M, Piva G (2009). On the occurrence of aflatoxin M1 in milk and dairy products. *Food Chem Toxicol*, 47(5): 984-91.
3. Battacone G, Nudda A, Palomba M, Pascale M, Nicolussi P, Pulina G (2005). Transfer of aflatoxin B1 from feed to milk and from milk to curd and whey in dairy sheep fed artificially contaminated concentrates. *J Dairy Sci*, 88 (9): 3063-9.
4. Stoloff L, Van Egmond HP, Parks DL (1991). Rationales for the establishment of limits and regulations for mycotoxins. *Food Addit Contam*, 8: 213-22.
5. Liu Y, Wu F (2010). Global burden of aflatoxin-induced hepatocellular carcinoma: a risk assessment. *Environ Health Perspect*, 118 (6): 818-24.
6. Dragacci S, Gleizes E, Fremy JM, Candlish AA (1992). Use of immunoaffinity chro-

- matography as a purification step for the determination of aflatoxin M1 in cheeses. *Food Addit Contam*, 12(1): 59-65.
7. Kocabas CN, Sekerel BE (2003). Does systemic exposure to aflatoxin B(1) cause allergic sensitization? *Allergy*, 58(4): 363-5.
 8. Park DL (2002). Effect of processing on aflatoxin. *Adv Exp Med Biol*, 504: 173-9.
 9. Food and Agriculture Organization, Worldwide regulations for mycotoxins in food and feed in 2003, Food and Agriculture Organization, Rome (2004) (*FAO Food and Nutrition Paper*, 81).
 10. Zheng H, Yunliang Z, Lianjun L, Zengxuan C, Yiping R, Yongjiang W (2010). An ultra-high-performance liquid chromatography-tandem mass spectrometry method for simultaneous determination of aflatoxins B1, B2, G1, G2, M1 and M2 in traditional Chinese medicines. *Analytica Chimica Acta*, 664: 165-71.
 11. Karimi G, Hassanzade M, Teimuri M, Nazari F, Nili A (2007). Aflatoxin M1 contamination in pasteurized milk in Mashhad Iran. *Iranian J Pharm Sci*, 3: 153-6.
 12. Alborzi S, Pourabbas B, Rashidi M, Astaneh B (2006). Aflatoxin M1 contamination in pasteurized milk in Shiraz. *Food Control*, 17: 582-4.
 13. Tajkarimi M, Shojaee Aliabadi F, Salah Nejad M, Pursoltani H, Motallebi AA, Mahdavi H (2007). Seasonal study of aflatoxin M1 contamination in milk in five regions in Iran. *Int J Food Microbiol*, 116(3): 346-9.
 14. Kirk GD, Bah E, Montesano R (2006). Molecular epidemiology of human liver cancer: insights into etiology, pathogenesis and prevention from The Gambia, West Africa. *Carcinogenesis*, 27(10): 2070-82.