

# Molecular Analysis of Iranian Patients with Duchenne/Becker Muscular Dystrophies

*S Kheradmand kia<sup>1</sup>, \*DD Farhud<sup>1</sup>, S Zeinali<sup>2</sup>, AR Mowjoodi<sup>2</sup>, H Najmabadi<sup>3</sup>,  
F Pourfarzad<sup>3</sup>, P Derakhshandeh<sup>1</sup>*

<sup>1</sup>*Dept. of Human Genetics & Anthropology, School of Public Health, Tehran University of Medical Sciences, Iran*

<sup>2</sup>*Dept. of Biotechnology, Pasteur Institute of Iran, Tehran, Iran*

<sup>3</sup>*University of Welfare and Rehabilitation, Tehran, Iran*

---

---

## Abstract

Duchenne Muscular Dystrophy (DMD) and the milder allelic Becker Muscular Dystrophy (BMD) are X-linked disorders. Both DMD & BMD result from heterogenous mutation in the dystrophin gene and in about 65% of the cases one or more exons of the gene are deleted or duplicated. One third of cases arise from new mutation and the rest are familial. To analyze the prevalence of deletion in Iranian patients, a deletion screening was performed on group 18 exons of dystrophin gene. Deletions were detected in 56.8% of patients. Seventy four percent of deleted exons were located in the major hot spot region, whereas 26% were in the minor hot spot one. The most frequently deleted exons were exons 50, 48 & 47 16.2%, 16.2% & 12% respectively. No deletion was detected in exon 43. The intragenic RFLP analysis (pERT87-15/BamHI & pERT87-8/TaqI) were carried out on DNA samples obtained from 22 Iranian unrelated families (196 males & females) showing DMD & BMD clinical symptoms, that 45% of them had informative patterns. The percentage of heterozygosity was 22.75% for BamHI intragenic RFLP, and 22.75% for TaqI intragenic RFLP.

**Keywords:** Duchenne, Becker, Muscular dystrophies, DMD/BMD, Dystrophin gene, Deletion, Iran

---

---

## Introduction

Duchenne and Becker Muscular Dystrophies (DMD/BMD) are allelic neuromuscular disorders caused by mutations to the dystrophin gene at Xp21 (1). Because of the extremely large size of this gene (nearly 2400 kb) the majority of mutations are intragenic deletions (~65%) or duplication (~5%) (2). In both DMD and BMD, partial deletions and duplications cluster in two recombination hot spots, one proximal at the 5' end of the gene, comprising exons 2-20 (30%), and one more distal, composing exons 44-53 (70%) (3). The severe Duchenne form is caused by the absence of detectable dystrophin, which, at the DNA level, is usually a result of mutations that alter translational reading frame leading to production of a truncated and the presumably unstable dystrophin muscle. On the other hand,

the milder BMD result from internal deletions or duplications that do not disrupt the reading frame, so partially functional portion can still be produced (4,5). Two multiplex polymerase chain reactions based assays encompassing 18 of 79 exons of the dystrophin gene have been developed and detect approximately 98% of deletions in DMD/BMD patients (6, 7, 8). Because of the severity of DMD and the lack of any effective treatment, it is imperative that an accurate carrier test be available to identify those female relatives, including mothers, of an affected individual who are at risk of having an affected son. Linkage analysis using marker flanking and within the dystrophin gene can predict with high accuracy whether the woman has inherited the same chromosome a common maternal relative as has an affected male.

## Materials and Methods

A total of 389 blood samples were collected from 106 Iranian DMD/BMD individual families with 207 affected males. The age of affected males ranged from 3 to 33 years. Ascertainment of clinical disease was made by the criteria described previously (9, 10), therefore 88 unrelated families with 182 patients were selected. Only one affected member per family was considered. For carrier detection with RFLP method we had 38 unrelated families with familial history that 22 of them were analyzed (they had at least one candidate for carrier detection). Genomic DNA was isolated from peripheral blood lymphocytes by the standard protocol (11). Screening for deletions was carried out essentially as described (7, 8). For Multiplex

analysis 18 exons (Pm, 3, 4, 6, 8, 12,13, 17, 19, 43, 44, 45, 47, 48, 50, 51, 52, 60) were amplified in three multiplex reactions (Set I, II, III). After amplification PCR products were electrophoresed in 3% agarose gel. The gel was stained with ethidium bromide and the bands were visualized by UV illumination (Fig. 1) demonstrates a multiplex reaction in 3 panels. Two intragenic RFLP systems (PERT 87-8/TaqI, PERT87-15/ BamHI) were used for familial cases (11). After amplification, 20µl of PCR products were digested with 5 units of either BamHI or TaqI for at least 1 h at 37°C or 60°C, as appropriate, electrophoresed on 3% agarose gel and visualized by ethidium bromide staining or 8% polyacrylamide gel detected by silver staining (Fig. 2).



**Fig. 1:** DMD/BMD deletion screening for 20 exons of dystrophine gene  
Set I: Pm Exons 3, 43, 50, 13, 6, 47, 60, 52 (Lane 1 exon 50 and 47 deleted)  
Set II: Exons 49, 51, 12, 4, 46 (Lane 1 exon 49 and 46 deleted)  
Set III: Exons 45, 48, 19, 17, 8, 44 (Lane 1 exon 19,17,8 deleted)  
M: DNA size marker  
Lane: 1-6 different patients  
N: Normal



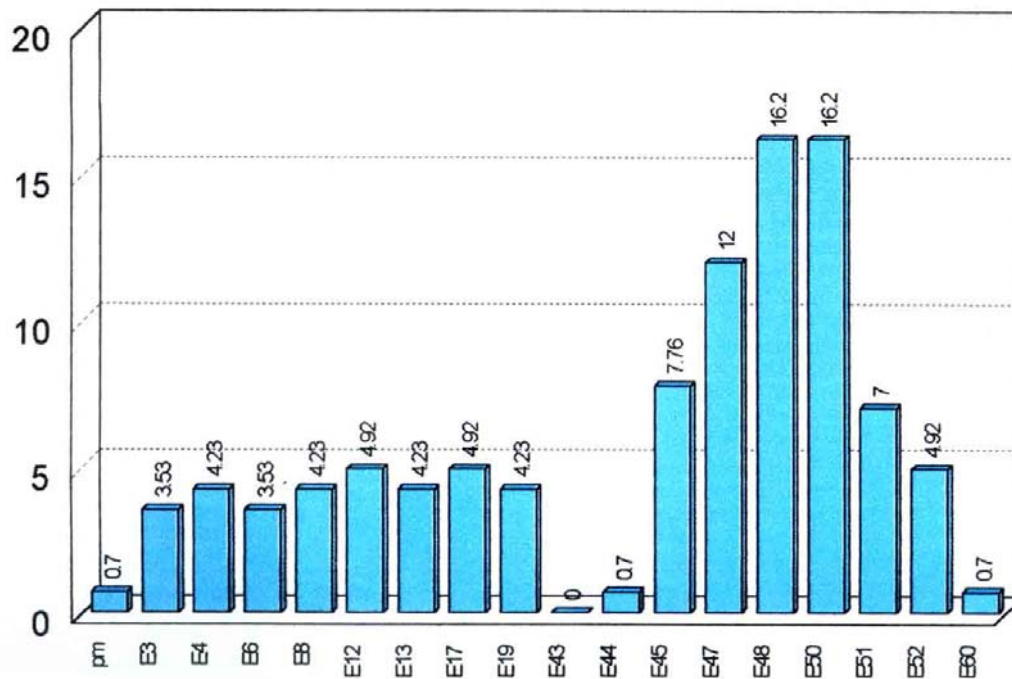
**Fig. 2:** BamHI and TaqI, RFLP gel electrophoresis BamHI was informative for this family

## Results

Examples of deletions detected by multiplex sets I, II, III are shown in Figure 1. Deletions were detected in 56.8% of patients. Seventy four percent of deleted exons were located in the major hot spot region, whereas 26% were in the minor hot spot region. The most frequently deleted exons were exons 50, 48 & 47 (16.2%, 16.2% and 12% respectively) (Fig. 3). No deletion was detected in exon 43. 36% of patients had only one exon deletion and 64% of them had more than one exon deletion. The number of exons deleted in a given child varied

from 1 to 9. The intragenic RFLP analysis (pERT87-15/BamHI & pERT87-8/TaqI) were carried out on DNA samples obtained from 22 Iranian unrelated families with familial history of DMD/BMD (196 males & females), that 45.5% of them had informative patterns. The percentage of heterozygosity was 22.75% for BamHI intragenic RFLP and same result for TaqI intragenic RFLP. Only two families were informative for both systems. Frequency of positive and negative allele in BamHI were 75%, 25% respectively and for TaqI site were 63.9% , 36.1% respectively.





**Fig. 3:** Frequency of dystrophin gene deletion among 18 exons in this study

## Discussion

This study showed 56.8% deletion rate in DMD/BMD patients. Fig. 4 compares the detection rates from North American & European studies varied from 55 to 70% (12,13,14), while in Asia it was lower, being 40-43% in Japan (15), 45-50% in China (16) and 55% in Thailand (17). In Turkish DMD patients, deletions were found in 52-59% (18, 19). But in Arab patients, deletions were found in 86% (20). In Greece, deletions were in 63% cases (21). The lowest rate of deletion mutations in DMD patients has been reported from the Philippines (33%) (22) and Israel (37%) (23). Amongst the Europeans, the lowest deletion rate is reported from Germany,

Hungary, Czechoslovakia and Spain (39-45%) (24). In Mexico DMD patients, deletions were detected in 52.5% cases (25). In 74% cases, deleted exons were restricted to the distal hot spot (exons 44-60), in 26% cases they were found in the proximal hot spot (exons 3-19), which is compatible with other reported studies. We have used PCR for rapid analysis of intragenic RFLP to permit both carriers and prenatal diagnosis in the majority of familial cases. Over 45% of all mothers at risk of carrying DMD/BMD are expected to be heterozygous for at least one of the two intragenic RFLP, which have studied. These results are compatible with other studies (11).

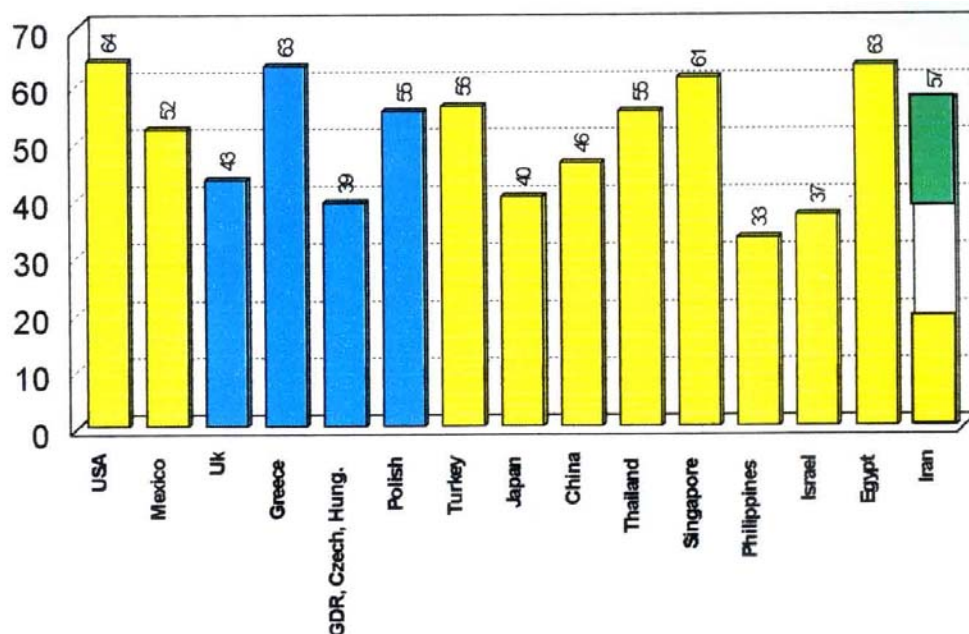


Fig. 4: Frequency of dystrophin gene deletion in the world

### Acknowledgements

This study was supported by grants from the Pasture Institute of Iran. We thank Farhud medical Genetic Clinic, for providing the patients material. Dr Najl Rahim, Dr Sigaroodi and Dr Shafaghati are gratefully acknowledged for clinical contribution. Special thanks are due to two technical assistants of Biotechnology Dept. of Pasture Institute, Zohre Arjang and Zahra Moghaddam.

### References

- Hoffman EP, Kunkel LM (1989). Dystrophin abnormalities in Duchenne and Becker muscular dystrophy. *Neuron*. 2: 1019- 29.
- Koeing M, Beggs AH, Meyer M, Scherpf S, Heinderichs K, Bettecken T, Meng G, Muller CR, Lindlof M, Kaariamien H, Chapelle A de la, Kiuru A, Saavonataus M-L, Gilgenkrantz H, Recan D, Chelly J, Kaplan J-C, Covone AE, Archidiacono N, Romeo G, Liechi – Gallatti S, Schneider V, Braga S, Moser H, Darras BT, Murphy P, Francke U, Chen JD, Morgan G, Denton M, Greeneberg CR, Wrogemann K, Blondon LAJ, Paassen HMBvan, Ommen GJB van, Kunkle LM (1989). The molecular basis for Duchenne versus Becker muscular dystrophy: correlation of severity with type of deletion. *Am J Hum Genet*. 45: 498 – 506.
- Hoffman Ep, Kunkle LM, Angelini C, Clarke A, Johnson M, Harris JB. (1989). Improved diagnosis of Becker muscular dystrophy via dystrophin testing. *Neurology*. 39: 1011-17.
- Den Dunnen JT, Grootscholten PM, Bakker E, Blonden LAJ, Ginjaar HB, Wapenaar MC, Paassen HMBvan, Broeckhoven C, Pearson PL, Ommen GJB (1989). Topography of the Duchenne muscular dystrophy (DMD) gene: FIGE and cDNA analysis of 194 cases reveals 115 deletions and 13 duplications. *Am J Hum Genet*. 45: 835 – 47.

5. Monaco AP, Bertelson CJ, Liechti-Gallati S, Moser H, Kunkle LM (1988). An explanation for the phenotypic differences between patients bearing partial deletion of DMD locus. *Genomics*. 2: 90-5.
6. Chamberlin JS, Gibs RA, Ranier JE, Casaki CT (1989). Deletion screening of the Duchenne muscular dystrophy locus via multiplex DNA amplification. *Nucleic Acids Res*. 16: 11141- 11156.
7. Chamberlin JS, Gibs RA, Ranier JE, Casaki CT (1992). Deletion screening gene: Analysis of Duchenne and Becker muscular dystrophy patients in Quebec. *Hum Genet*. 89: 419- 24.
8. Beggs AH, Koeing M, Boyce FM, Kunkle LM (1990). Detection of 98% of DMD/BMD gene deletions by polymerase chain reaction. *Hum Genet*. 86: 45-8.
9. Miller G, Wessel H (1992). Diagnosis of dystrophinopathies: Review for the clinician. *Pediatr Neural*. 9: 3-9.
10. Emmery AEH. (1988). *Duchenne muscular dystrophy*, ed 2. New York, Oxford University Press.
11. Roberts RG, Cole CG, Hart KA, Bobrow M, Bentley DR (1989). Rapid carrier and prenatal diagnosis of Duchenne and Becker muscular dystrophy. *Nucleic Acids Res*. 17: 811.
12. Liechti – Gallati S, Koeing M, Kunkel LM, Frey D, Botshauer E, Schneider V, Braga S, Moser H (1989). Molecular deletion patterns in Duchenne and Becker muscular dystrophy. *Hum Genet*. 81: 343- 48.
13. Simard LR, Gingras F, Delvoeye N, Vanasse M, Melancon SB, Labuda D (1991). Deletions in the Duchenne and Becker muscular dystrophy gene in the Chinese. *Am J Med Genet*. 89: 419- 424.
14. Hodgson S, Hart K, Abss S, Heckmatt J, Rodillo E, Bobrow M, Dubowitz V (1992). Correlation of clinical and deletion data in Duchenne and Becker muscular dystrophy. *J Med Genet*. 89: 343- 48.
15. Immoto N, Arinami T, Hamono K, Matsumura K, Yamada H, Hagaguchi H, Takito H (1993). Topography pattern of rearrangement of the dystrophin gene in Japanese Duchenne muscular dystrophy. *Hum Genet*. 92: 533- 36.
16. Soong BW, Tsai TF, Su TS., Takumio T, Masamura T, Koga J, Nakajima T, Masummura T, Koga J, Nakamura H. (1992). Amplification of ten deletion rich exons of the dystrophin gene by polymerase chain reaction shows deletions in 36 of 90 Japanese families with Duchenne and Becker muscular dystrophy. *Am J Med Genet*. 42: 453 – 457.
17. Mutirangura A, Norapusuntun T, Srivuthana S, et al. (1992). Multiplex PCR to detect the dystrophin gene deletion in Thai patients. *J Trop Pediatr*. 38: 224- 27.
18. Battaloglu E, Telatar M, Deymeer F, Serdargolu P, Kuseyri F, Ozdemir C, Apak M, Tolun A (1992). DNA analysis in Turkish Duchenne and Becker muscular dystrophy Families. *Hum Genet*. 89: 635 – 39.
19. Kuseyri GN, Topalogulu H, Apak M, Kirdar B (1993). Screening of deletions and RFLP analysis in Turkish Duchenne and Becker muscular dystrophy families by PCR. *Clin Genet*. 43: 261 –66.
20. Haidar MZ, Bastaki L, Habib Y, Moosa A (1998). Screening 25 dystrophin gene exons for deletions in Arab children with Duchenne and Becker muscular dystrophy. *Hum Hered*. 48: 61- 6.
21. Florentin L, Mavrou A, Kekou K, Metaxotou G (1995). Deletion patterns of Duchenne and Becker muscular dystrophines in Greece. *J Med Genet*. 32: 48- 51.

22. Cutingeo CM, Padilla C, Takenaka Y, Yamasaki Y, Matus M, Nishio H. (1994). More deletions in the 5' region than in the central region of the dystrophin gene were identified among Filipino Duchenne and Becker muscular dystrophy patients. *Am J Med Genet.* 49: 369 – 73.
23. Shomrat R, Gluck E, Legum C, Shiloh Y (1994). Relatively low proportion of dystrophin gene deletions in Israel Duchenne and Becker muscular dystrophy patients. *Am J Med Genet.* 49: 379 – 83.
24. Speer A, Kraft U, Grade K, Coutelle C, Wulff K, Wehnert M, Herrmann FH, Kadasi L, Kunert E, Muller U, Foster C, Wolf C, Szibor R (1990). Deletion analysis of DMD/BMD families from German Democratic republic and selected regions of Czechoslovakia and Hungary. *J Med Genet.* 27: 679 – 82.
25. Carol – Vazquez R, Arenas D, Cisneros B, Penaloza L, Kofman S, Salamanca F, Montanez C (1993). Analysis of dystrophin gene deletions in patients from Mexican population with Duchenne and Becker muscular dystrophy. *Arch Med Res.* 24: 1-6.