

SUBTYPE DETERMINATION OF PHOSPHOGLUCOMUTASE (PGM) AND GROUP - SPECIFIC COMPONENT (Gc) BY ISOELECTRIC FOCUSING AMONG SOME IRANIAN POPULATIONS

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Abstract

A total of 781 serum and 827 red cell samples from four populations of Iran (Kashan, Ardestan, Astara and Ardabil) were sub-typed by isoelectric focusing for group-specific component and red cell enzyme Phosphoglucomutase, locus one, respectively. Heterogeneity test revealed no significant differences among the populations studied regarding both PGM1 and Gc systems.

Introduction

Phosphoglucomutase (PGM) is an important enzyme in carbohydrate metabolism since it catalyzes the interconversion of glucose 1 - phosphate and glucose 6 - phosphate in the presence of catalytic amounts of the cofactor glucose 1, 6 - diphosphate.

Investigations of phosphoglucomutase locus one (PGM1) phenotypes by isoelectric focusing have shown four common alleles at this locus (2,3,11,16,20). Various reports (3,4,24) have shown that these alleles are determined by one locus with Mendelian inheritance.

The Group-Specific component (Gc) polymorphism previously detected by starch gel electrophoresis (4,10,15,18) was characterized by two common alleles Gc¹ and Gc² in all human populations. Later on, isoelectric focusing technique (5,6) revealed different electrophoretic patterns. These patterns can be explained by the existence of three autosomal codominant Gc^{1P}, Gc^{1S} and Gc² genes controlling the synthesis of the protein bands revealed by electrophoretic mobilities. When the first data on gene frequencies became

available, the allele Gc² turned out to have low frequencies in areas of high aridity.

This finding was explained when the function of the Gc proteins was discovered (7). They are carrier proteins of vitamin D.

A more recent survey suggests a relationship between sunshine and the Gc polymorphism, in the aboriginal habitats of the world populations, high frequencies of Gc² were found in most of the populations who had been living for a long time in areas with low sunlight intensity. This geographical distribution suggests a selective advantage due to a more efficient transport of vitamin D, especially if the supply is limited (12).

Materials and methods

To study phosphoglucomutase locus 1 and group - specific component systems, blood samples were collected from four Iranian populations located in north and central parts of the country. Blood was drawn into syringes, transferred to vials and transported at the earliest possible time at wet ice temperature, to Manchester, England, for analysis. Serum and red cells were separated, haemolysates prepared and stored at -20 °C until tested. 827 red cell haemolysates were tested for PGM1 sub - types. Acid starch gel electrophoresis was performed (14). Isozymes were visualized by the staining technique (19). 781 serum samples were sub - typed for group specific component (6).

Results and discussion

The populations studied and their observed and expected numbers of PGM₁, and Gc phenotypes and their gene frequencies are given in tables 1 and 2 respectively. X² was not significant and showed lack of deviation from Hardy - Weinberg equilibrium in either systems in all the populations studied. Test of heterogeneity among the population studied for both PGM₁ and Gc systems showed lack of heterogeneity (X² = 18.3 for PGM₁ and X² = 24.4 for Gc), indicating that populations do not differ significantly for subtypes studies.

The highest gene frequency for Gc^{1S} was observed in Kashan and the lowest in Ardabil populations, for Gc^{1P} the highest value occurred in Ardestan

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and the lowest in Ardabil. The highest value for Gc² was observed in Ardabil and the lowest in Kashan.

In PGM₁ system, the highest gene frequency for PGM₁¹⁺ occurred in Astara and the lowest in Ardabil, for PGM₁¹⁻ the highest occurred in Ardabil and the lowest in Astara.

For PGM₁²⁺, the highest was in Ardabil and the lowest in Astara. For PGM₁²⁻ the highest was in Kashan and the lowest in Astara.

With regard to PGM₁¹⁺ allele values, around 60-70%, found in the present investigation, showed affinity to Europeans frequencies. Frequencies of PGM₁¹⁺ is low and close to previous report from Iranian Zoroastrians (14) and the combined frequency of the PGM₁¹⁺ and PGM₁¹⁻ alleles ranging from 0.747 to 0.764 is very similar to that of migrant Parsis and Iranis studied in India (21,22), and is within the range for the Iranian populations reported previously (8,17).

Regarding Gc system, Gc^{1F} allele frequency in the present study is much lower than that in Iraq and Israel (13), and well fits to the European range of 12 - 20%. This is true for Gc^{1S} frequency too.

The sum of the allele frequencies of Gc^{1S} and Gc^{1F} in the populations studied is from 0.673 (Ardabil) to 0.808 (Kashan), which is close to the previous reports (1,14,23).

Nei's genetic distance estimated for PGM1 subtypes (Table 3) showed Astara population to have less similarity to other populations followed by Ardabil.

The same values for Gc subtypes showed less similarity of Ardabil population with others, followed by Astara. Hence Ardabil and Astara populations are well separated from other two populations with regard to PGM1 and Gc subtypes (Table 4).

The results indicate the necessity of further investigations on other populations of Iran in order to reach a sound conclusion, however present study well indicates differences among the populations.

High frequencies of Gc², seems to exist among populations of Ardabil and Astara, where the sunlight intensity in contrast with the areas of Ardestan and Kashan is low (12). In conclusion the different geographical distribution of Gc² in the present study may be an association between Gc gene

frequencies and diet, incident ultraviolet - a function of cloud cover latitude and altitude - or, possibly skin color. We emphasize that in the absence of specific supportive evidence of a biochemical or physiological nature, any environmental correlation with Gc phenotype can at least be only suggestive, and further studies of large samples from different populations of Iran will certainly be profitable.

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Table 1- Observed and expected numbers of PGM1 and Gc subtypes in four Iranian populations.

| System/ Phenotype | Populations | | | | | | | |
|----------------------|-------------|------|----------|-------|--------|------|---------|-------|
| | Kashan | | Ardestan | | Astara | | Ardabil | |
| | Obs. | Exp. | Obs. | Exp. | Obs. | Exp. | Obs. | Exp. |
| PGM1 | | | | | | | | |
| 1 + 1 | 89 | 82.6 | 104 | 100.1 | 84 | 81 | 122 | 115.7 |
| 1 + 1 - | 19 | 19.6 | 20 | 21.7 | 15 | 9.5 | 26 | 29.7 |
| 1 - 1 - | 2 | 1.2 | 3 | 1.2 | 1 | 0.7 | 5 | 2.0 |
| 1 + 2 + | 32 | 39.3 | 43 | 48.1 | 29 | 36.3 | 53 | 60.8 |
| 1 - 2 + | 2 | 4.6 | 4 | 5.2 | 6 | 3.3 | 8 | 7.8 |
| 1 + 2 - | 15 | 19.5 | 24 | 22.3 | 19 | 16.7 | 27 | 26.0 |
| 1 - 2 - | 4 | 2.3 | 2 | 2.4 | 0 | 1.5 | 1 | 3.3 |
| 2 + 2 + | 9 | 4.7 | 11 | 5.8 | 7 | 4.1 | 10 | 8.0 |
| 2 + 2 - | 6 | 4.6 | 7 | 5.4 | 3 | 3.8 | 11 | 6.8 |
| 2 - 2 - | 2 | 1.2 | 0 | 1.2 | 1 | 0.9 | 1 | 1.5 |
| Total | 180 | | 218 | | 165 | | 264 | |
| Gc | | | | | | | | |
| 1S - 1S | 79 | 79.3 | 67 | 66.5 | 64 | 62.4 | 72 | 69.4 |
| 1S - 1F | 37 | 37.3 | 39 | 39.2 | 27 | 27.1 | 30 | 33.6 |
| 1F - 1F | 5 | 4.4 | 6 | 5.7 | 3 | 2.9 | 5 | 4.0 |
| 2 - 1S | 47 | 46.5 | 55 | 55.6 | 48 | 51.1 | 82 | 83.6 |
| 2 - 1F | 10 | 10.9 | 16 | 16.34 | 11 | 11.1 | 22 | 20.2 |
| 2 - 2 | 7 | 6.8 | 12 | 11.6 | 12 | 10.4 | 25 | 25.2 |
| Total | 185 | | 195 | | 165 | | 236 | |

Table 2- Gene frequencies of PGM1 and Gc systems in four Iranian populations.

| System/ Phenotype | Populations | | | |
|--------------------------------|-------------|----------|--------|---------|
| | Kashan | Ardestan | Astara | Ardabil |
| PGM ₁ | | | | |
| PGM ₁ ^{1*} | 0.6775 | 0.6775 | 0.70 | 0.662 |
| PGM ₁ ^{1*} | 0.0805 | 0.0735 | 0.0641 | 0.085 |
| PGM ₁ ^{2*} | 0.161 | 0.163 | 0.157 | 0.174 |
| PGM ₁ ^{2*} | 0.0980 | 0.0755 | 0.0725 | 0.0744 |
| Gc | | | | |
| Gc ^{1S} | 0.6540 | 0.5840 | 0.6115 | 0.5425 |
| Gc ^{1F} | 0.1540 | 0.1720 | 0.1335 | 0.1310 |
| Gc ² | 0.1920 | 0.2440 | 0.2515 | 0.3265 |

Table 3- NEI'S genetic distance for PGM subtypes

| | Kashan | Ardestan | Astara | Ardebil |
|----------|-----------|-----------|-----------|-----------|
| Kashan | 0.0000000 | | | |
| Ardestan | 0.0001211 | 0.0000000 | | |
| Astara | 0.1531892 | 0.1599088 | 0.0000000 | |
| Ardebil | 0.0004464 | 0.0004403 | 0.1455595 | 0.0000000 |

Table 4 - NEI'S genetic distance for Gc subtypes

| | Kashan | Ardestan | Astara | Ardebil |
|----------|-----------|-----------|-----------|-----------|
| Kashan | 0.0000000 | | | |
| Ardestan | 0.0032187 | 0.0000000 | | |
| Astara | 0.0076603 | 0.0032831 | 0.0000000 | |
| Ardebil | 0.0271980 | 0.0142900 | 0.0062587 | 0.0000000 |

References

- 1- Bajatzadeh, M. and Walter, H. (1968): Serum Protein polymorphisms in Iran. *Human genetik*, 6, 40 - 54.
- 2- Bark, J.E., Harris, M.J. and Firth, M. (1976): Typing of the common PGM variants using isoelectric focusing, a new interpretation of the PGM system. *Journal of the Forensic Science Society*, 16, 115 - 120.
- 3- Bissobort, S., Ritter, H. and Kompf, J. (1978): PGM1 Subtyping by means of acid starch gel electrophoresis. *Hum. Genet*, 45, 175 - 177.
- 4- Carter, N.D., West, C.M., Emes, E., Parkins, B. and Marshal, W.H. (1979): PGM polymorphism detected by isoelectric focusing gene frequencies, evaluation and linkage. *Ann. Hum. Bio.* 6, 221-230.
- 5- Constans, N. and Viau, M. (1977): Group-Specific component evidence of two subtypes of the Gc. *Gene. Science*, 198. 1070-1071.
- 6- Constans, J., Viau, M., Cleve, H., Quilici, J.C. and Palisson, M.J. (1978): Analysis of the Gc polymorphism in human populations by isoelectric focusing on polyacrylamid gels. *Hum. Genet.* 41, 53 - 60.
- 7- Daiger, S.P., Schanfield, M.S. and Cavali - Sforza, L.L. (1976): Gc protein Bind Vitamin D. and 25 - Hydroxy Vitamin D. *Proc. Nat. Acad. Sci. USA.* 72, 6, 2076 - 2080.
- 8- Farhud, D.D., Ananthakrishnan, R., Walter, M. and Loser, J. (1973): Electrophoretic investigations of some red cell enzymes in Iran. *Human Heredity*, 23.263 - 266.
- 9- Kirk, R.L., Cleve, H. and Bearn, A.G (1963): The distribution of the Group-Specific Component Gc in Selected population in south and south east Asia and Oceania, *Acta. Genet. Bascl.* 13, 140 - 149.
- 10- Kirk, R.L., Cleve, H. and Bearn, A.g. (1963): The distribution of the Gc types in sera from Australian aborigins. *Am. J. Phgs. Anthrop.*
- 11- Kuhl, P., Schmidtman, U. and Spielmann, W. (1977): Evidence for two additional common alleles at the PGM1 locus. *Hum. Genet.* 35, 219 - 223.
- 12- Mourant, A.E., Tills, D. and Domaniewska - Sobczak, K. (1976): Sunshine and the geographical distribution of the allele of the Gc system of plasma proteins. *Hum. Genet.* 33, 307- 314.
- 13- Papiha, S.S., Roberts, D.F., White, I., Chahal, S.M. and Asefi, A.J. (1982): Population genetics of group specific component (Gc) and PGM Studied by Isoelectric focusing *Am. G. Phys. Ant.* 18, 76-79.
- 14- Papiha, S.S., Seyedna, S.Y. and Sunderland, E. (1982): PGM and Gc Isoelectric focusing subtypes among Zoroastrians of Iran. *Ann. Hum. Biol* 9, 6, 571 - 574.
- 15- Parker, A.G., Cleve, H. and Bearn, A.G. (1963): Determination of phenotypes in the human Group - Specific component (Gc) system by starch gel electrophoresis. *Am. Hum. Genet* 21, 230 - 236.
- 16- Santachiara - Benereceti, A.S., Razani, G.N. and Antonini, G. (1981): Subtyping of human red cell PGM1 Polymorphism, a third PGM₁¹ allele common among twapggmies from north Rwanda.
- 17- Sawhney, K.S. (1975): Genetic Polymorphisms in selected populations in south west and South Asia. Ph.D thesis, Durham University, U.K.
- 18- Schanfield, M.S., Eugene Giles, and Gershowitz, H. (1975): Genetic studies in the Markham valley, Northeastern Papua new Guinea. *Am. J. Phys. Anthrop*, 42, 1-8.
- 19- Spencer, N., Hopkinson, D.A. and Harris, H. (1964): PGM Polymorphism in man. *Nature*, 204, 742-745.
- 20- Sufton, J.G. and Burgess, R. (1978): Genetic evidience for four common alleles at the PGM1 detected by isoelectric focusing. *Vox Sanguinis.* 34, 97-103.
- 21- Undevia, J.V., Blake, N.M., Kirk, R.L. and Mcdermeid, E.M. (1972): The distribution of some enzyme group system among Parsis and Iranis in Bombay. *Hum. Hered.* 22, 274-282.
- 22- Undevia, J.V., Kirk, R.L. and Me dermid, E.M. (1973): Serum Protein System among Parsis and Iranis in Bombay. *Hum. Hered.* 23, 492-498.
- 23- Walter, V.H. and Djahanshahi, M. (1983): Zur Haufigkeit der Serumgruppen in persien. *Homo*, 14, 70-76.
- 24- Welch, S.G., Swindlehurst, C.H., Mc Georger, I.A. and Williams, K. (1978): Isoelectric focusing of human red cell phosphoglucomutase. *Hum. Genet.* 43, 307-313.